



EUROPEAN MEDICINES AGENCY
SCIENCE MEDICINES HEALTH

22 April 2010
EMA/CHMP/37646/2009
Committee for Medicinal Products for Human Use (CHMP)

Guideline on the use of pharmacogenetic methodologies in the pharmacokinetic evaluation of medicinal products

Draft

Draft Agreed by Pharmacogenomics Working Party and EWP- PK	March 2010
Adoption by CHMP for release for consultation	22 April 2010
End of consultation (deadline for comments)	31 October 2010

This guideline replaces the Reflection Paper on the use of Pharmacogenetics in the Pharmacokinetic Evaluation of Medicinal Products (EMA/128517/2006)

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Keywords	<i>Pharmacogenetics, pharmacokinetics, clinical development</i>
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1 Executive summary

2 This guideline addresses the influence of pharmacogenetics on drug pharmacokinetics, encompassing
3 considerations and requirements for the design and conduct of investigations during drug development.
4 In particular, guidance is given regarding studies required and recommended at different phases of
5 drug development to ensure satisfactory efficacy and safety in pharmacogenetic subpopulations that
6 have variable systemic exposure of active substances.

7 1. Introduction (background)

8 The pharmacokinetics of many medicinal products is prone to interindividual variability, which is
9 caused by several factors such as gender, age, weight, renal and hepatic function, and genetics. In
10 recent years, a rapid development in our understanding of the influence of genes on interindividual
11 differences in drug action has occurred. This development encompasses the area of pharmacogenomics,
12 including pharmacogenetics, where interindividual variability in genes influencing or predicting the
13 outcome of drug treatment (e.g., genes encoding drug transporters, drug metabolising enzymes, drug
14 targets, biomarker genes) is studied in relation to efficacy of drug treatment and adverse drug
15 reactions. A great deal of this interindividual variability is caused by genetic polymorphism, i.e. the
16 occurrence in the same population of multiple allelic states. Genetic variations are demonstrated by the
17 identification of Single Nucleotide Polymorphisms (SNPs), insertions/deletions and variation in gene
18 copy number (copy number variation, CNV).

19 With respect to pharmacokinetics, the highest level of polymorphism is found in genes involved in drug
20 metabolism; phase I metabolism of approximately 40% of clinically used drugs is due to polymorphic
21 enzymes. Currently, the most important polymorphic enzymes are the cytochrome P450 enzymes such
22 as CYP2C9, CYP2C19 and CYP2D6. Subjects who have extensive and poor metabolising capacity for
23 these enzymes are present in the general population. For CYP2D6, besides the poor metaboliser
24 phenotype, the ultrarapid metaboliser phenotype is relevant as well. With respect to phase II enzymes,
25 the genetic variability of UDP-glucuronosyltransferases, *N*-acetyltransferase-2 and some
26 methyltransferases has been linked to interindividual pharmacokinetic variability. The metabolising
27 enzymes account for 80% of the drugs which currently include pharmacogenetic data in their labelling.

28 Among the major clinically relevant issues is pharmacogenetic variability causing increased or
29 decreased metabolism of the parent drug and the subsequent formation of active or toxic substances.
30 Decreased metabolism can cause too high levels of the parent drug and adverse drug reactions.
31 Elevated drug metabolism can cause loss of response or, in case of prodrug activation, too high levels
32 of the bioactive compound.

33 The interindividual genetically linked differences in pharmacokinetics may cause, clinically, very
34 relevant alterations in drug action. Optimal efficacy is dependent on appropriate dosing, often based on
35 the specific genotype. Thus, the effective dose may vary greatly due to increased or decreased drug
36 elimination rate. For instance, due to CYP2D6 polymorphism, the rate of hepatic metabolism of drugs
37 which are substrates for this enzyme can vary 1000-fold between individuals. Among many
38 antidepressants and antipsychotics, the plasma levels of the drug at the same dosage often vary 5-20-
39 fold. A 9-fold higher risk of suicide has been reported among ultrarapid metabolisers of CYP2D6 and
40 there are also many reports of increased frequency of adverse drug reactions among subjects with the
41 poor metaboliser phenotype, due to increased systemic exposure of the parent drug. Furthermore,
42 increased side effects after treatment with analgesic drugs, which are activated by CYP2D6, are seen
43 among ultrarapid metabolisers. The efficacy of prodrugs which are activated by polymorphic enzymes
44 varies depending on the pharmacogenetics of the patients. An example of this is clopidogrel, for which
45 an increased frequency in serious side effects due to excessive prodrug activation has been seen in
46 patients with increased formation of the active metabolite and a corresponding lack of effect in
47 subjects without the appropriate enzyme. Dosing of some important anticoagulants is dependent on
48 the CYP2C9 genotype of the patient and the platelet inhibition action of some drugs is dependent on
49 the CYP2C19 genotype. Overall, it can be estimated that 20-25 % of the efficacy of all drug treatment
50 is significantly affected by interindividual differences in genes encoding drug metabolising enzymes.

51 In recent years, journal articles have been published describing specific polymorphisms in drug
52 transporters and their possible effect on the efficacy and safety of medicinal products. However, in the
53 majority of cases the influence of transporter polymorphism on drug pharmacokinetics has not yet
54 been clarified. The effect of transporter polymorphism on drug pharmacokinetics is thought to be of
55 less importance, or is still unknown, compared with polymorphic enzymes. This is partly due to the fact
56 that the role of specific transporters *in vivo* is difficult to quantify due to the lack of specific inhibitors

57 and the polymorphism seen in the transporters is often substrate specific in its effects. The possibility
58 of transporter polymorphism as a cause of altered pharmacokinetics must, however, always be
59 considered in all phases of drug development. It is anticipated that this area will expand a great deal in
60 the near future, as knowledge of the role of drug transporters is rapidly developing.

61 At present, an increasing proportion of lead compounds selected for further development are
62 metabolised by enzymes or transported by transporter proteins upon which the impact of
63 pharmacogenetics is unknown. New technologies, such as rapid genome sequencing, whole genome
64 wide association studies (GWAS) and targeted absorption, distribution, metabolism and excretion
65 (ADME) gene SNP/CNV analyses, are expected to have an integral role in clinical drug development in
66 the future.

67 Until now, it has been difficult to transfer knowledge of the effect of polymorphism into specific
68 recommendations in affected genetic subpopulations¹. In this respect, genetic subpopulations have
69 been treated differently than other subpopulations or circumstances in which the exposure of active or
70 toxic substances is increased. The aim of including pharmacokinetics-related pharmacogenetics in drug
71 development is to evaluate whether exposure in genetic subpopulations is different to such an extent
72 that this would require a change in the posology or treatment recommendation of the drug for the
73 specific subpopulation. This document, therefore, aims to clarify the studies needed to investigate
74 these issues.

75 **2. Scope**

76 The aim of this guideline is to clarify the requirements related to the use of pharmacogenetics in the
77 pharmacokinetic evaluation of medicinal products.

78 The following issues are discussed in this guideline:

- 79 • In which situations and at what stage(s) in the clinical development program should
80 pharmacogenetic/pharmacokinetic studies be performed.
- 81 • Recommendations regarding pharmacogenetic/pharmacokinetic studies investigating the
82 effects of polymorphisms at ADME level (enzymes and transporters), including study design,
83 selection of subjects, and sampling.
- 84 • Evaluation of the clinical impact of genetic differences on pharmacokinetic parameters and
85 recommendations on further studies to support the posology/treatment recommendations for
86 genetic subpopulations.
- 87 • Possible consequences of genetic differences on pharmacokinetic parameters for treatment
88 recommendations and labelling.
- 89 • Special considerations related to drug-drug interactions and the effect of impaired or immature
90 organ functions on pharmacogenetics.

91 **3. Legal basis**

92 This guideline applies to Marketing Authorisation Applications for new medicines for human use
93 submitted in accordance with Article 8(3) of the Directive 2001/83/EC, as amended. This guideline
94 should be read in conjunction with the Introduction and general principles paragraph (4) and Part I,
95 Module 5 of the Annex I to Directive 2001/83, as amended, and all other relevant information included
96 in current and future EU and ICH guidelines and regulations especially:

- 97 • Pharmacokinetic studies in man (Notice to applicants, Vol 3C, C3a, 1987).
- 98 • Guideline on reporting the results of population pharmacokinetic analyses
99 (CHMP/EWP/185990/06).
- 100 • The investigation of drug interactions (CPMP/EWP/560/95).
- 101 • Guideline on the investigation of bioequivalence (CPMP/EWP/QWP/1401/98 Rev.1).
- 102 • Guideline on the role of pharmacokinetics in the development of medicinal products in the
103 paediatric population (EMA/CHMP/EWP/147013/2004).

¹ The term "genetic subpopulation" may include both the qualifiers of the polymorphisms, e.g. poor metaboliser, as well as the indication of the concerned allele, e.g., CYP 2D6*4.

- 104 • Guideline on the evaluation of the pharmacokinetics of medicinal products in patients with
105 impaired hepatic function (CPMP/EWP/2339/02).
- 106 • Note for guidance on the evaluation of the pharmacokinetics of medicinal products in patients
107 with impaired renal function (CHMP/EWP/225/02).
- 108 • Position paper on terminology in Pharmacogenetics (EMA/CPMP/3070/01).
- 109 • Rules governing medicinal products in the European Union Volume 2C Notice to applicants; A
110 guideline on summary of product characteristics (SmPC) September 2009.
- 111 • ICH Topic E15. Note for Guidance on definitions for Genomic biomarkers, pharmacogenomics,
112 pharmacogenetics, genomic data and sample coding categories
113 (EMA/CHMP/ICH/437986/2006).
- 114 • ICH Topic E16. Note for Guidance on genomic biomarkers related to drug response: context,
115 structure and format of qualification submissions. (EMA/CHMP/ICH/380636/2009).

116 **4. Situations and stage in development where the effect of** 117 **pharmacogenetics on pharmacokinetics should be studied**

118 **4.1. General recommendations**

119 Pharmacogenetic variants often influence drug pharmacodynamics but also the absorption, distribution,
120 metabolism and excretion of a drug. Furthermore, pharmacogenetics may also influence the formation
121 and elimination of metabolites and this should be remembered if there are metabolites that may affect
122 the efficacy and/or safety of the drug. Genotypes leading to absent, decreased or increased enzyme or
123 transport protein activity should be considered.

124 Studies of the effect of pharmacogenetics on the pharmacokinetics of an active substance (parent
125 and/or active metabolites) and its implications for efficacy and safety are generally **required** during
126 development:

- 127 a) If *in vitro* studies and/or *in vivo* studies indicate an important role for a functionally
128 polymorphic enzyme in the metabolism of the drug, *or*
- 129 b) If *in vitro* studies and/or *in vivo* studies indicate an important role for a functionally
130 polymorphic enzyme in the formation or elimination of a pharmacologically active or toxic
131 metabolite, *or*
- 132 c) If *in vivo* studies indicate that a transport protein known to be subject to clinically relevant
133 polymorphism has a significant effect on the systemic or target organ exposure of
134 pharmacologically active drug or metabolites.

135 Studies on the effect of pharmacogenetics on the pharmacokinetics of an active substance and its
136 implications for efficacy and safety are generally **recommended** during development:

- 137 a) if available *in vitro* data indicate that a polymorphic enzyme or transporter contributes to the
138 pharmacokinetics of the active substances but the quantitative role is unknown, *or*
- 139 b) if there is high interindividual pharmacokinetic variability, or there are pharmacokinetic outliers
140 with several-fold higher or markedly lower exposure to the active substances, which gives rise
141 to clinical efficacy and safety concerns based on the existing knowledge, *or*
- 142 c) if safety concerns are observed which may be related to genetic differences in systemic or
143 target organ exposure, *or*
- 144 d) if major differences in pharmacokinetics are observed in different ethnic groups.

145 If a studied polymorphism of an enzyme or transporter does not affect the functional performance of
146 the protein, genotyping² is not considered to be necessary during the clinical development program.
147 The same is true if the results of pharmacokinetic studies clearly show that the impact of
148 pharmacogenetics is not clinically relevant based on pre-specified, well supported target exposure
149 limits.

150 Still, in all clinical phases of development, prospective sampling of DNA for genotype analyses is
151 recommended, even when there are no obvious indications of a relevant genetic influence on
152 pharmacokinetics. This recommendation is due to the fact that unknown polymorphic sites of

² In this guideline the term genotyping is used. However, phenotyping by, for example, catalytic assays may also be an acceptable approach

153 importance can be identified later and that unknown but important metabolic pathways in which a
154 polymorphism might have an effect, may be identified at later stages in the programme.

155 **4.2. Integrating pharmacogenetic effects on pharmacokinetics in drug** 156 **development**

157 Below, recommendations are made on how to implement pharmacogenetics during the different
158 phases of clinical development, starting with the *in vitro* studies conducted before studying the
159 medicinal product in man.

160 **4.2.1. In vitro studies prior to human exposure.**

161 Prior to Phase I, human *in vitro* metabolism studies should be performed. Such studies should
162 preferably include identification of the enzymes catalysing the *in vitro* metabolism and also the
163 identification and characterisation of metabolites formed, enabling pharmacological activity screening
164 of these metabolites. It should be remembered that polymorphic enzymes can participate in the
165 formation and elimination of pharmacologically active metabolites of the drug, including toxic
166 metabolites. Even when these pathways quantitatively represent a minor pathway for drug elimination,
167 they may be highly significant in terms of safety or efficacy. Based on these data, the involvement of
168 known functionally polymorphic enzymes in the elimination of the parent compound and/or the
169 formation and elimination of active metabolites should be determined. As *in vitro* studies are not
170 always quantitatively predictive of the *in vivo* situation, the enzyme involvement needs to be confirmed
171 *in vivo*.

172 The involvement of transporters may be indicated by *in vitro* data obtained prior to the start of Phase I.
173 However, it is presently very difficult to make quantitative predictions of the *in vivo* contribution of
174 transporters. The *in vivo* importance of a transporter may be demonstrated by preclinical *in vivo*
175 information or information on similar substances.

176 **4.2.2. First time in man studies**

177 The possibility of genetic influence should be considered early in the Phase I program. When the *in*
178 *vitro* data indicate that involvement of a known functionally polymorphic enzyme cannot be excluded,
179 it is advised to genotype the first time in man study population. If the available *in vitro* data strongly
180 indicate major involvement of a polymorphic enzyme in the metabolism and/or formation of
181 pharmacologically active or toxic substances, subjects with a genotype predicted to result in markedly
182 increased exposure of active or toxic substances should, preferably, only be allowed to enter in the
183 first time in man study at doses several-fold lower than the doses expected to be safe in extensive
184 metabolisers.

185 Presently, knowledge of transporter proteins is not mature enough to estimate the potential for
186 significant involvement of the transporter *in vivo* based on *in vitro* data. Unless there are other
187 indications of significant transporter involvement *in vivo*, early genotyping is not required on the basis
188 of *in vitro* data. Still, prospective storing of samples in order to allow eventual pharmacogenetic
189 analysis is recommended.

190 If future knowledge of drug transporters expands to such extent that certain *in vitro* data on
191 transporters may be considered predictive of the clinical situation, the same protocol as described for
192 metabolising enzymes may also be appropriate for polymorphisms in drug transporter encoding genes.

193 **4.2.3. Phase I**

194 When based on available *in vitro* or preliminary clinical data, genotype is predicted or known to
195 markedly affect the pharmacokinetics of pharmacologically active compounds, genotyping is
196 recommended in as many of the Phase I studies as possible in order to increase the amount of data
197 that will support the recommendations for use in the genetic subpopulation(s).

198 In Phase I, the relative contribution of the identified polymorphic enzyme to total body clearance
199 should be estimated. If feasible, it is recommended to investigate this in a conventional
200 pharmacokinetic study with targeted inclusion of subjects of certain genotypes (see section 5.1). If this
201 is not feasible, but potent and sufficiently specific *in vivo* inhibitors of the enzyme are available, the
202 effect of the polymorphism should be estimated using the results of an interaction study with such an

203 inhibitor. The effect of the identified polymorphism on the maximum tolerated dose should also be
204 determined on the basis of these studies.

205 If a marked effect of polymorphism is confirmed, it is recommended, if feasible, to expand the clinical
206 Phase 1 program and also evaluate the interaction potential, as well as the consequences of impaired
207 organ function and age in the genetic subpopulations (see sections 8.1 and 8.2). Furthermore, dose-
208 linearity in poor metabolisers at relevant doses should be investigated. If pharmacokinetics are linear
209 in the general population, these data may be extrapolated to populations with increased protein
210 activity. However, if pharmacokinetics are nonlinear in extensive metabolisers, it is recommended to
211 investigate dose-proportionality in ultrarapid metabolisers, in order to provide satisfactory background
212 knowledge for the future studies. This evaluation should preferably be done before starting Phase II
213 and the results of this evaluation should be taken into consideration in the study protocol.

214 **4.2.4. Phase II (dose finding, exploratory)**

215 If Phase I studies strongly indicate that pharmacogenetics may influence the pharmacokinetics of a
216 drug to a clinically relevant extent, this should be considered in the design of the Phase II studies. It is
217 then required to genotype the included patients. Genotype based dosing or exclusion of certain
218 genotypes should be considered. The aim should be to minimise variability in exposure of
219 pharmacologically active substances for each dose-level studied and to optimise target dose selection
220 in the Phase III studies. The effect of the known polymorphism on pharmacokinetics, safety and
221 efficacy in the populations of altered genotype should be clarified. Dose recommendations for genetic
222 subpopulations should be worked out, based on, for instance, the pharmacokinetic characteristics and
223 pharmacodynamics of the new active substance. The expected level of evidence for a genotype-based
224 dose adjustment is comparable with that required for dose adjustment based on other intrinsic and
225 extrinsic factors affecting pharmacokinetics, like weight, age, renal and hepatic impairment or drug-
226 drug interactions. If active substance exposure in Phase III is not "normalised" through genotype
227 based dosing, dose titration or, less desirably, through genotype based exclusion of patients, sufficient
228 data needs to be collected on the consequences of the altered exposure on the clinical efficacy and
229 safety of the drug (see section 7).

230 **4.2.5. Phase III (confirmatory)**

231 If available data indicate that there is a significant difference in drug exposure in the pharmacogenetic
232 subpopulation, all patients included in phase III studies should be genotyped. Depending on the likely
233 consequences of the polymorphism for efficacy or safety, there are several ways that this knowledge
234 could be influence the design of Phase III studies:

235 a) No genotype-specific treatment (if the available data indicate that the difference in exposure may
236 not be clinically relevant, so there are no major safety or efficacy concerns related to the difference in
237 exposure).

238 b) Normalising the exposure of active substances through genotype based dosing.

239 c) Dose titration regardless of genotype (in case a suitable marker exists).

240 d) Exclusion of patients of a specific genotype (i.e., exclude patients at risk, if it is not possible to
241 normalise the exposure).

242 In case the exposure of active substance(s) is not normalised through genotype-based dosing, the
243 conclusion of similar efficacy and safety at the exposure obtained in the subpopulation needs to be
244 supported by sufficient clinical data obtained at these exposure levels to allow these conclusions. This
245 may included both Phase II and Phase III data.

246 The final aim of the clinical development program should be to obtain a clear dosing or treatment
247 recommendation, yielding effective and safe treatment in the genetic subpopulations.

248 **4.3. *Involvement of relevant polymorphic enzymes and proteins identified*** 249 ***in the course of the clinical development program***

250 In Section 4.2 of this guideline the ideal situation has been described, where the potential effect of
251 pharmacogenetics is detected early in drug development. In situations where there are no predictive
252 indications of pharmacogenetic effects on the pharmacokinetics of the drug when initiating the clinical
253 part of the development program of a new medicinal product, the acquired pharmacokinetic, clinical

254 efficacy and safety information at a certain stage in the drug development may indicate the need for
255 investigations of the pharmacogenetic impact on drug or metabolite exposure. This situation may occur:

256 a) when a previously unknown or sparsely studied enzyme or transporter is found to be involved in the
257 metabolism or transport of the medicinal product that is being developed,

258 b) if the enzyme or transporter is known but there was no prior knowledge regarding polymorphisms of
259 the gene,

260 c) if knowledge of the importance of an enzyme or transporter to the pharmacokinetics of a drug is
261 gained at a late stage in the development program,

262 d) if high interindividual pharmacokinetic variability, or marked outliers in drug exposure are observed,

263 e) if there are safety or efficacy concerns observed which may originate from pharmacogenetic
264 differences in local tissue or systemic drug exposure, or,

265 f) if there are major differences in the pharmacokinetics of the medicinal product between different
266 ethnic groups.

267 See section 4.1 for situations in which pharmacogenetic investigations are required or recommended.

268 If data become available, during the clinical development program or during pharmacovigilance
269 monitoring, demonstrating the involvement of a polymorphic enzyme or transporter as a cause for
270 important inter-individual variations in pharmacokinetics, it is recommended to, as far as possible,
271 estimate the relative contribution of the metabolism or transport pathway in question to the
272 bioavailability and/or clearance of the medicinal product *in vivo*. If needed, it is recommended that
273 relevant *in vitro* studies are carried out in which the direct influence of the polymorphic variant enzyme
274 is characterised. See also section 4.2.

275 If the genetic locus associated with the identified pharmacokinetic variability influencing clinical or
276 safety aspects is unknown, it is advisable to carry out analysis (such as phenotype-genotype
277 associations) of genes potentially responsible for the variation. More thorough investigations using
278 modern techniques such as GWAS, genome sequencing, or array based SNP/CNV investigations are
279 recommended to identify the background (see sections 5.2 and 5.3).

280 Conclusions from a retrospective analysis carried out in response to emerging data may be acceptable
281 if mechanistically supported by available pharmacokinetic information. In this case, DNA should
282 preferably be available from a large proportion of patients. If a new genetic association is discovered in
283 a retrospective analysis, complementary studies, such as *in vitro* studies or pharmacokinetic studies
284 investigating the mechanism and pharmacokinetic consequences of this finding, are recommended.

285 In specific cases it may be appropriate to contact the European Medicines Agency to discuss the issue
286 during a pharmacogenetic briefing meeting or a Scientific Advice meeting.

287 **5. Study design and methodology**

288 ***5.1. Conventional pharmacokinetic analysis and population*** 289 ***pharmacokinetic analysis***

290 The requirements during drug development with regard to monitoring pharmacogenetics related to
291 pharmacokinetics depend on the stage of the clinical development plan and how strong the evidence is
292 for a clinically relevant effect of pharmacogenetics on drug exposure. *In silico* Physiologically Based
293 Pharmacokinetic (PBPK) modelling and simulation may be helpful when optimising the design of *in vivo*
294 pharmacokinetic studies.

295 The effect of polymorphism on the exposure of pharmacologically active substances should, if possible,
296 be investigated in a pharmacokinetic study of conventional, frequent blood sampling, design. A Phase I
297 study of reduced design, including the extremes of genotypes (e.g., extensive metabolisers vs. poor
298 and/or ultra-rapid metabolisers) is usually performed as a basis for the evaluation of the
299 pharmacogenetic effect on active substance exposure. However, if a likely clinically relevant effect is
300 found, the consequences of having an intermediate protein activity should also be investigated. The
301 study may be of single-dose design. If the pharmacokinetics are nonlinear and a dose range is
302 recommended, the highest and lowest dose of the range should be covered (see section 4.2.3). If an
303 effect of genotype is observed in the single-dose study, a multiple dose study is needed unless dose

304 linearity has been shown at the concentrations obtained in carriers of the low activity (e.g. poor
305 metabolisers) genotypes and the pharmacokinetics of the drug is independent of time.

306 The conventional pharmacokinetic study should, in principle, include enough subjects for a likely
307 clinically relevant difference in exposure to be detected between the included genotypes. However, if
308 homozygotes for the allele(s) giving rise either to the most reduced or highest activity are difficult to
309 recruit as the allele is very rare, as many carriers of the rare extreme genotype as possible should be
310 included. In this case, the study may include a larger number of heterozygote carriers or gene variant
311 carriers having an intermediate protein activity. This may allow a preliminary estimation of the
312 consequences of this polymorphism in subjects who are homozygous for the variant. In this respect,
313 available data from other drugs may be helpful to support the relation between genotype and
314 phenotype.

315 Population pharmacokinetic analysis of datasets, with or without the inclusion of rich data sets, could
316 be used to confirm the effect of pharmacogenetics on active substance exposure. It may also be used
317 as a hypothesis-generating tool where the effect of new or unexpected polymorphisms may be
318 indicated. The study population should include a satisfactory number of patients of each genotype, and
319 enough samples per patient to obtain valid estimates. Power calculations and sampling optimisation
320 should be done before the initiation of the study in order to ensure a sufficient population size. If the
321 data and analysis is of high quality and the study sufficiently powered, the data may be used to
322 support labelling.

323 However, if a need for pharmacogenetics based dose adjustments has been identified, this should,
324 generally, be supported by a conventional pharmacokinetic study, as in such cases a more precise
325 estimation of the genotype effect is needed. Population pharmacokinetic analysis of sparse data from
326 Phase III may be used as supportive data indicating that a genotype-based dosing or treatment
327 recommendation applied in Phase III has normalised drug exposure in the patient population.

328 **5.2. Genotyping methods and choice of alleles**

329 At present, a rapid development is taking place with respect to analytical methods available for allele
330 specific genotyping, such as real time polymerase chain reaction (PCR), oligonucleotide ligation assay
331 (OLA), TaqMan PCR, SNP/CNV arrays, pyrosequencing, genomic sequencing, etc. In the case of
332 targeted genotyping, the method should first be validated utilizing well characterised standard samples
333 carrying the mutation in question, preferably both in the heterozygous and homozygous states. When
334 using PCR techniques it is important to continuously analyse blank samples only containing
335 water/buffer in order to exclude contamination reactions. The polymorphisms studied should, in
336 principle, be those where the genetic alteration causes changes in gene function or be strongly
337 associated with documented phenotypic variants. To obtain reliable data it is important to include a
338 large enough number of samples to allow for rare alleles, in order to provide enough power for reliable
339 statistical calculations. The genetic analyses should, preferably, also encompass assays for CNVs, a
340 type of polymorphism of increasing importance.

341 In cases where high interindividual variability in pharmacokinetics is observed without any likely
342 hypothesis regarding the genetic origin, commercial arrays for analyses of the 10,000- 30,000 most
343 relevant SNPs and CNVs in ADME genes are now available and could provide relevant results regarding
344 the genetic origin of the pharmacokinetic variation. In cases where a significant association is made to
345 the pharmacokinetic variation in question, it is important to analyse the true functional polymorphic
346 site, which might be in linkage disequilibrium to the SNPs/CNVs present on the chip. Modelling and
347 simulation methods can help in analysing the data and designing further studies as pharmacogenetic
348 variants can be incorporated in the models as variables of interest.

349 **5.3. Genome wide association studies**

350 In cases where the relevant hypotheses for the genetic basis of interindividual variation in
351 pharmacokinetics cannot be placed, genome wide association studies (GWAS) now offer an alternative
352 method for identification of the true locus of importance. Today SNP/CNV arrays monitoring more than
353 1.1 million genetic alternations in the human genome are commercially available. Such arrays are now
354 more routinely used for identification of disease causing genes and have also been useful in the
355 identification of genetic variations e.g. in the MHC Class II region responsible for increased sensitivity
356 to adverse drug reactions caused by certain drugs. Note should be taken of emerging GWAS

357 knowledge in relevant public databases³. In case no explanation for observed variability is found,
358 GWAS studies to reveal interindividual differences in pharmacokinetics and pharmacodynamics of
359 drugs are recommended in the clinical development of drugs. Recent published results show that
360 GWAS has been of use in the identification of variable alleles responsible for altered response or dosing
361 toxicity of several different drugs. The significance of the linkage between the phenotype and the
362 mutation/CNV must reach a high statistical level and results should, preferably, be obtained from a
363 second independent cohort. Furthermore, the SNP associated with the phenotype may not be the true
364 causative mutation. In these cases it is recommended to sequence the loci in the neighbouring regions,
365 analyse the SNPs linked to the initial SNP detected and analyse the data for significance level by taking
366 the new SNP(s) into consideration. If the significance level is increased, the functional aspects of the
367 mutation have to be analysed. The new mutations discovered in these studies should, preferably, be
368 validated *in vitro* and *in vivo*. The functional consequences can be identified using cellular transfection
369 of variant plasmids. The *in vivo* importance can be evaluated by retrospective stratification of
370 previously characterised data with respect to the occurrence of the mutations and by prospective
371 studies using patients selected by genotype.

372 **6. Presentation of study results**

373 **6.1. Conventional pharmacokinetic studies**

374 Individual data on pharmacokinetic parameters, like AUC, C_{max} , t_{max} , CL/F or CL and F, and $t_{1/2}$ in
375 relation to genotypes should be listed. Standard descriptive statistics for each genetic subpopulation,
376 including mean, standard deviation and range should be provided for the pharmacokinetic parameters.
377 The parameters representing drug exposure (e.g. C_{max} and AUC) could be presented for separate
378 subgroups as box-whiskers-plots. The plots should include the individual data points either overlaid or
379 next to the boxes.

380 The effect of pharmacogenetics on pharmacokinetics of the investigational drug should be calculated
381 and the relative difference in relevant pharmacokinetic parameters presented. The 90% confidence
382 interval for the genotype effect should be presented.

383 If the pharmacokinetics of active metabolites has been investigated, the data should be presented in a
384 similar way for the metabolites. If suitable, active moiety, i.e. the sum of the unbound exposure of
385 pharmacological equivalents, should be presented in addition to the effects of the separate substances.
386 However, this estimation is only correct if the distributions of parent drug and metabolite to the target
387 site(s) are similar. The validity of this assumption should be discussed and, if possible, the calculations
388 could be modified by the metabolite to parent target organ distribution ratio.

389 **6.2. Population pharmacokinetic analysis**

390 Reference is made to the *Guideline on reporting the results of population pharmacokinetic analyses*
391 (CHMP/EWP/185990/06).

392 **6.3. Genotyping methods and Genome wide association studies**

393 With respect to the presentation of genotyping methodologies and outcomes, reference is made to the
394 *Note for Guidance on genomic biomarkers related to drug response: context, structure and format of*
395 *qualification submissions*. ICH Topic E16 (EMAE/CHMP/ICH/380636/2009).

396 **6.4. Phase II and III studies**

397 If appropriate, the same applies here as described for pharmacokinetic studies. It is acknowledged that
398 in Phase II and III studies, full pharmacokinetic data will not always be available. Still, available
399 pharmacokinetic or population pharmacokinetic data in relation to genotypes should be listed, and
400 standard descriptive statistics for each genetic subpopulation, including mean, standard deviation and
401 range should be provided for the pharmacokinetic parameters.

³, e.g., the HuGE Navigator (<http://hugenavigator.net/>), the NIH Database of Genotype and Phenotype (<http://www.ncbi.nlm.nih.gov/gap>) or the Catalog of Genome-Wide Association Studies (<http://www.genome.gov/GWASudies/>)

402 With respect to clinical data obtained with respect to pharmacogenetics, reference is made to the *Note*
403 *for Guidance on genomic biomarkers related to drug response: context, structure and format of*
404 *qualification submissions*. ICH Topic E16 (EMAE/CHMP/ICH/380636/2009).

405 **7. Evaluation of the clinical consequences of genetic** 406 **differences and translation into treatment recommendations**

407 The clinical consequences of observed differences in drug exposure in genetic subpopulations depend
408 on several factors, such as:

- 409 • the magnitude of the difference in exposure caused by the polymorphism,
- 410 • the relationship between pharmacokinetics and pharmacodynamics of the medicinal
411 product,
- 412 • the relationship between drug exposure and clinical effects/adverse effects,
- 413 • severity of the possible adverse events and clinical consequences of loss of efficacy.

414 Dosing recommendations should ensure that patients receive drug treatment which is effective and
415 safe. Preferably, a genetic effect should be compensated by adjusting the dose of the drug to achieve
416 an exposure which is shown to be effective and safe. Either genotype-base dosing is applied or the
417 dose individualisation is performed through individual dose titration based on Therapeutic Drug
418 Monitoring (TDM), efficacy or adverse events.

419 When a polymorphism in a metabolising enzyme or transporter leads to a difference in exposure which
420 may alter efficacy or safety, the expected level of evidence for showing that the proposed treatment
421 recommendation is suitable for the subpopulation is comparable with that required for effects of other
422 intrinsic or extrinsic factors affecting pharmacokinetics. The evaluation of clinical consequences should
423 be based on information available on the relationship between exposure and efficacy/safety. If possible,
424 a well justified target range for relevant exposure parameters should be presented for the
425 investigational drug specifying what change in exposure would justify a posology adjustment. The
426 target range is the range of drug exposure for which satisfactory clinical efficacy and safety has been
427 shown. If the target range is based on drug exposure in patients and the pharmacogenetic effect was
428 investigated in healthy volunteers, potential differences between the pharmacokinetics of patients and
429 healthy subjects need to be considered. The observed exposure (box-whiskers plots including
430 individual data) should be analysed with respect to target criteria taking into account the frequency of
431 patients with lower as well as higher exposure than the target range and the clinical consequences of
432 these deviations. Unless the applicant convincingly shows that the exposure obtained in the genetic
433 subpopulation is effective and safe, the proposed dosing recommendation in the genetic subpopulation
434 should “normalise” drug exposure. Efficacy and safety in the absence of normalised drug exposure
435 should, preferably, be based on Phase II and III data in a sufficient number of individuals exposed to
436 the same active substance exposure. Knowledge gained from similar drugs at increased exposure is
437 also supportive.

438 If the parent drug is pharmacologically active and there are *in vivo* relevant active metabolites, the
439 exposure of these metabolites should be taken into account when proposing dose adjustments. When
440 relevant, the active moiety can be used to develop dose adjustment (see also section 6.1). However,
441 increased exposure of the separate substances must also be considered. The exposure of all relevant
442 active substances should, as far as possible, be within a well tolerated range after dose adjustment.

443 In case the genetic subpopulation is too small to allow thorough clinical investigation of proposed dose
444 adjustment, it is recommended that the resulting individual exposure parameters obtained with the
445 proposed treatment recommendation are estimated and the safety and efficacy expected of the
446 resulting exposure is evaluated as described in section 5.1.

447 In general, clinical impact and treatment recommendations should be evaluated separately for the
448 homozygous and heterozygous population, since in most cases clinical consequences are expected to
449 differ between these subgroups.

450 If the proposed dose-adjustment is based on C_{min} as a surrogate for AUC, it should be taken into
451 account that the relation between C_{min} and AUC may be altered, if the systemic elimination of the drug
452 is changed.

453 **8. Special pharmacogenetics considerations with respect to**
454 **drug-drug interactions, impaired/immature organ functions**
455 **and age**

456 **8.1. Drug interactions**

457 Polymorphisms in metabolising enzymes can not only affect the exposure of the pharmacologically
458 active substances, but also impact on the drug-drug interaction profile. Therefore, the possible effects
459 of polymorphisms on the interaction profile should be evaluated.

460 If a major elimination pathway is absent or very diminished in a subpopulation (e.g., in poor
461 metabolisers), other elimination pathways will be of increased importance. The consequences of
462 inhibition of these alternative pathways should be determined and reflected in study protocols as well
463 as treatment recommendations, if the drug will be used in the subpopulation. In case a drug
464 interaction study in the subpopulation is not possible, a worst case estimation should be made based
465 on the available *in vivo* knowledge of the quantitative contribution of separate enzymes to drug
466 elimination. If the subpopulation is not identified (e.g., through genotyping) before combining the
467 interacting drugs, the general treatment recommendations should be suitable for the subpopulation.
468 The potential need to adjust any treatment recommendation related to the effects of other drugs on
469 the pharmacokinetics of the drug under investigation, or vice versa, should be considered.

470 As a general rule, genotyping of the population included in a drug-drug interaction study is
471 recommended when pharmacogenetics are expected to affect the pharmacokinetics of any of the drugs
472 included in the study. Depending on the aim of the study, a list of inclusion or exclusion criteria may
473 contain specific genotypes.

474 **8.2. Impaired or immature organ function and age**

475 The consequences of impaired organ function may be different in genetically different subpopulations.
476 This applies if the main elimination pathway in the genetic subpopulation is markedly affected by the
477 impaired organ function. The exposure of active substances resulting from impaired organ function in
478 the genetic subpopulation should be estimated and the clinical consequences discussed based on the
479 available safety data. Additional information should be included in the SPC to reflect this.

480 The enzymes and transport proteins involved in the pharmacokinetics of a drug substance may be
481 quantitatively and qualitatively different in the very young paediatric patients than in adults as a
482 consequence of developmental gene expression. Such differences are mainly expected in newborn
483 infants, infants and toddlers (0-2 year-old children).

484 If a significant impact of a genetic polymorphism on the pharmacokinetics of a drug substance has
485 been established in adults, the potential consequences in the paediatric population should be further
486 elaborated and taken into account during drug development. Likewise, it should be considered that in
487 the very elderly patients, pharmacogenetic related differences in metabolism could occur and have a
488 different impact on efficacy and safety, due to impaired functionality of system/organs.

489 **9. Specific issues related to treatment recommendations**
490 **based on genetically determined differences in exposure**

491 Labelling text referring to genotype testing may be: 1) for information purposes only, 2) recommended
492 or 3) mandatory. This will depend on the strength of the data available and on the efficacy and safety
493 consequences expected.

494 **9.1. Dose recommendations**

495 Different routes for dose adjustment can be applied:

496 *1) Dose titration*

497 Differences in exposure in genetic subpopulations can be managed by dose-titration in all patients
498 based on safety and/or efficacy markers, or on TDM. If this approach is chosen, the applicant needs to

499 show that the titration schedule is suitable for the specific subpopulation(s) as well as for the general
500 patient population (see section 7).

501 *2) Optional gene based dosing*

502 If the expected consequences of having a different exposure of active substances due to genetic
503 polymorphism are undesirable, for instance, adverse events affecting quality of life, but not sufficiently
504 severe for mandatory genotyping to be required, the option of improving the benefit/risk ratio via
505 genotyping prior to treatment should be mentioned in the SPC. In this case, suitable treatment
506 recommendations should be given for the genetic subpopulation.

507 When knowledge is available that genetics might aid in individual dose optimisation, an approach such
508 as safety-based titration can be enriched with a genetic component (e.g. with algorithms for thiopurine
509 S-methyltransferase (TPMT) variants and 6-mercaptopurine dosing in acute lymphatic leukaemia).

510 *3) Dosing based on genotype*

511 If a dose titration is not satisfactory or feasible and the safety or efficacy consequences of the
512 exposure difference in the subpopulation are considered to be a major concern, the genotype should
513 be carefully ascertained before initiation of therapy and appropriate dose adjustments should be
514 recommended for each relevant genetic subpopulation. If it is not possible to administer appropriate
515 doses with the available formulation strengths, a contraindication should be considered based on the
516 benefit-risk ratio of the treatment for the population concerned. The applicant is then encouraged to
517 develop suitable formulations to allow dose adjustment.

518 In both case 2 and 3, effort should be made to provide clear information and recommendations to the
519 prescriber. When relevant, recommendations shall be provided in Section 4.2. In most situations it is
520 sufficient to indicate the phenotypes (e.g. poor, extensive, ultrarapid metabolisers) in section 4.2, with
521 reference to section 5.2. In section 5.2, detailed information on the effect of different genotypes on
522 active substance exposure should be included and, if relevant, the effect on pharmacodynamics in
523 section 5.1 of the SPC. In case 2, available information on the difference in adverse events should be
524 presented in section 4.8.

525 **9.2. Other labelling consequences**

526 If a suitable dose can not be recommended based on available data, this should be reflected in the SPC,
527 e.g. as warnings, contra-indications, etc.

528 Well-documented functional polymorphisms that have not been studied because of their rare
529 appearance should be reflected in the SPC if they are likely to influence drug exposure to a clinically
530 relevant extent.

531 The frequencies of the alleles of interest in ethnic populations should be presented in the SPC section
532 5.2.

533 If genotyping is recommended, or optional, this should also be mentioned in the Product Information
534 Leaflet (PIL).